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Efficacy of Imipenem and Tigecycline in a Mouse Pneumonia Model Infected with Acinetobacter baumannii

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Abstract

Background: A. beumannii (ABAU) is an important clinical pathogen primarily associated with nosocomial pneumonia and/or bacteremia infections. Due to a confinued need for evaluating novel agents that treat drug-resistant ABAU infections, a mouse pneumonia model was developed that can evaluate antibiotic efficacy with both CPU (colony-forming unt) and survival

Introduction

Acinetobacter baumannii infections have become a major concern among healthcare facilifies worldwide, which is primarily the result of two important facts. The first fact is that both the prevalence and incidence of antibiofic-resistance among A. baumannii cinical isolates have significantly increased over the last two decades. The second fact is that A. baumannii can

significantly increased over the last two decades. The second fact is that A. Baumanni can survive for protonge periods within the nopilipati-enviroment, thus increasing the lighty hood of infectious spread among valenable patients. Additionally, A. Baumanni can easily coincise menulincul verialization and other devices, which is one of the reasons with hospital-acquired and the contraction of the contract A. baumannii. Here we describe the use of this model to evaluate the efficacy of Tigecycline

Methods and Materials

Minimum inhibitory concentrations (MICs): MICs were determined for Imipenem and Tigecycline against both A. baumannii clirical strains (UNT091-1 and UNT092-1) by using the microditution method as described by the Clinical and Laboratory Standards Institute (CLSI).

Mouse oneumonia model

Animals: Female, 5 - 6 wk old CD-1 mice (18 - 22 gm) were used according to the protocol approved by the UNTHSC-Institutional Animal Care and Use Committee.

Neutropenia: Niice were rendered neutropenic by intraperitoneally (i.p.) injecting cyclophosphamide at 150 mg/kg 4 days prior to infection, followed by 100 mg/kg on the day

before infection.

Incoculum proparation & infection: A. beumannii strains (UNT091-1 and UNT092-1) were cultured overnight on tryptic soy agar (TSA), and then suspended to the desired inoculum size for each strain. Anesthetized mice (ketamine at 40 mg/kg, xylazine at 6 mg/kg) were infected with the prepared inoculaby intransaci institlation.

he prepared inocula by intranasal institlation. ment. All doses of each antibiotic (mipnenm or Tigecycline) were administered by i.p. ion. For colony-forming unit (CFU) studies, mice were given a single dose of the nated antibiotic and specified dose amount (mg/kg) at 4 hours post-infection. For such ies, antibiotics were initially administered 4 hours post-infection, and then dosed twice daily

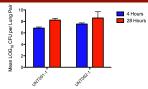
nints: For CELI studies, mice were euthanized at 4 (controls) and 28 hrs nost-infection Employmes. For Ore studies, increased, and then plated onto tryptic say agar (TRA) that their lungs were removed, homogenized, and then plated onto tryptic say agar (TRA) that contained activated charcoal. For survival studies were monitored for survival over 10 total days, which included a period of 7 days following the last antibiotic doos.

Panel 1: MIC Values for Imipenem and Tigecycline against A. baumannii Strains UNT091-1 and UNT092-1

	UNT091-1	UNT092-1
Imipenem	8 μg/mL	2 μg/mL
Tigecycline	2 μg/mL	2 μg/mL

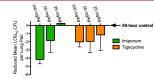
 MIC values determined by the microdilution method according to CLSI guidelines. A. baumannii reference strain ATCC19606 was included as a quality control for each MIC test (data not shown).

Panel 2: Lung CFUs of UNT091-1 and UNT092-1 at 4 and 28 Hours Post-Infection



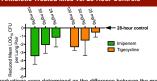
- CFU counts of lungs harvested at 4 & 28 hours post-infection for each strain.
- · Lungs were homogenized, serial diluted, and plated onto TSA+charcoal. • Error bars represent the standard deviation (SD) of the mean for each time point. (n=3)

Panel 3: Mean Log₁₀ CFU Reduction of <u>UNT091-1</u> in the Lungs of Antibiotic Treated Mice vs. 28-Hour Controls



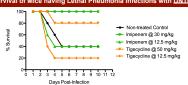
- Mean log₁₀ CFU reductions were determined as the difference between the mean CFU counts of untreated 28-hour controls vs. antibiotic treated groups.
- Indicated doses (x-axis) represent the amount intraperitoneally administered as a single dose 4hours after infection. Lungs were taken 28 hours after the final dose.
- Error bars represent the SD of the reduced mean CFUs for each group. (n=3)

Panel 4: Mean Log₁₀ CFU Reduction of <u>UNT092-1</u> in the Lungs of Antibiotic Treated Mice vs. 28-Hour Controls



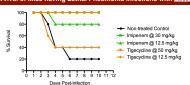
- Mean log of CFU reductions were determined as the difference between the mean CFU. counts of untreated 28-hour controls vs. antibiotic treated groups.
- Indicated doses (x-axis) represent the amount intraperitoneally administered as a single dose 4-hours after infection. Lungs were taken 28 hours after the final dose.
- Error bars represent the SD of the reduced mean CFUs for each group. (n=3)

Panel 5: % Survival of Mice having Lethal Pneumonia Infections with <u>UNT091-1</u>



- Percent survival of untreated and antibiotic treated mice for 10 days following infection with
- Indicated doses (legend) represent the amount intraperitoneally administered as a single dose 4-hours after infection, and then twice daily for 3 consecutive days.

Panel 6: % Survival of Mice having Lethal Pneumonia Infections with <u>UNT092-1</u>



- · Percent survival of untreated and antibiotic treated mice for 10 days following infection with
- Indicated doses (legend) represent the amount intraperitoneally administered as a single dose 4-hours after infection, and then twice daily for 3 consecutive days.

Summary and Conclusions

- Minimum inhibitory concentration (MIC) values indicate that both A. baumanni clinical strains, UNT091-1 and UNT092-1, were equally sensitive to Tigocytine (Panel 1). However, the MIC value for timipener against UNT091-1 was 4-fold higher than the one generated for UNT092-1, suggesting that does of timipenem may be less efficacious in mice with respiratory infections caused by UNT091-1.
- ➤ Lung-associated CFUs for strain UNT091-1 increased from 6.82 log_{so} at 4 Lung-associated CFUs for strain UNT091-1 increased from 6.82 (log, at 4 hours post-infection to 8.22 (log, at 28 hours, white the CFUs for strain UNT092-1 increased from 7.53 to 8.55 (log, over the same 28-hour period (Panel 2). The increase in lung CFUs over 28 hours for both strains indicated that a metaboliciatly active and growing infection was generated in mice, which implied that this model could be used to evaluate the efficacy of antibiotics against A. baumannii respiratory infections.
- Planels 3 8. 4) While single doses of Impenen at 50 and 100 mg/kg decreased lung CFUs to between 1.8 to 4.2 log, in LNT0611-1 infected animals, a single dose of Impenen at 50 and 50 mg/kg dot of declerely reduce total lung counts as compared to 28-hour controls. For mice infected with NT0621-1 single doses of Impenen at 25, 50, and 100 mg/kg reduced total lung CFUs to between 1.1 to 3.4 log, when compared to 28-hour controls except to 28-hour controls of 28-hour controls.
- CP as compared to the boal using course of 22-bits of commons.

 Planets 5.6 B) Multiple does of Imprement over 3.5 days at 30 mg/kg resulted in complete protection (100% survival) of mice that were leftably infected with the Park Boarmanit status, Imprement does at 12.5 mg/kg over 3.5 days resulted in the 40% survival of mice infected with protection of the common status of the survival of mice infected with NUTROS-1. Theopycline doses of 12.5 and 50 mg/kg over 3.5 days resulted in 20% to 80% of the cost survival of mice infected with NUTROS-1. The same doses of Tige-pc/laire but NUTROS-1 infected mice resulted in 40% to 100% of the animals surviving over the contrast of mice to 90% of 100% of the animals surviving over the contrast of 100% to 100%.

References

- 1)Allen DM & Hartman BJ. (1995). Acinetobacter species. In Principles and Practices of infectious Diseases, 4th edn (Madell, G. L., Jr. Douglas, R. G. & Bennett, J. E., Eds), 2009-13. Churchill Livingstone, New York. 2)Gordon NC, Wareham DW. Multi-resistant Acinetobacter Isuumannii:
- 2/Gordon NC, Wareham DW, Multi-resistant Anniechader baumanni: Mechanisms of Virulence and Resistance. Int J Antimicrobia Algents. 2010; 35(9): 219 228.
 2010; 35(9): 219 229.
 1900; 19
- Chemother. 2000. 45(4):493 501.
 5)Siegel JD, Rhinehart E, Jackson M, Chiarello L. Healthcare Infection Control Practices Advisory Committee. US Centers for Disease Control and Prevention. Accessed January 25, 2008.

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